



# JCIMPT

JOINT CENTER FOR INNOVATIVE MEMBRANE PROTEIN TECHNOLOGIES

## Cell Surface Expression Assay

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Version 1.0

### Summary:

This protocol is used by JCIMPT to determine the cell surface expression of FLAG tagged constructs. Production of structure-grade mammalian membrane proteins in substantial quantities has been hindered by a lack of methods for effectively profiling multiple constructs expression in higher eukaryotic systems such as insect or mammalian cells. To address this problem, a specialized small-scale eukaryotic expression platform by Thomson Instrument Company (Vertiga-IM) was developed and used in tandem with a Guava EasyCyte microcapillary 96-well cytometer to monitor cell density and health and evaluate membrane protein expression. The reference (Hanson et al. 2007) given below provides detailed description of work done to develop a construct profiling method that uses this protocol.

### Materials:

#### 1. Reagents:

- a. Cell strain expressing the desired construct.
- b. Positive control.
- c. Anti-FLAG M2 Monoclonal Antibody (SIGMA, catalog number F-3165).
- d. Zenon Alexa Fluor 488 Mouse IgG<sub>1</sub> Labeling Kit (Invitrogen, catalog number Z25002)

#### 2. Equipment:

- a. Guava EasyCyte flow cytometer ([Guava Technologies](#))

### Step by step methodology

- a. Prepare a 0.2 µg/µL antibody mix in Zenon Alexa Fluor 488. Incubate 5 minutes at room temperature. Dilute 1:10 with Tris Buffered Saline (TBS) containing 4% BSA (final concentration of antibody = 0.02 µg/µL). You will need 15 µL of the mix per sample.

*Sample calculation for 54 samples:*

- 54 samples x 15 µL antibody mix/sample = 810 µL total antibody mix
- Always make up a little extra, so prepare a total of 900 µL.
- 900 µL x 0.02 µg/µL antibody = 18 µg antibody



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- 18  $\mu\text{g}$  antibody / (10  $\mu\text{g}/\mu\text{L}$ ) (concentration of anti-FLAG antibody) = 1.8  $\mu\text{L}$  antibody
  - Mix 1.8  $\mu\text{L}$  antibody with 87  $\mu\text{L}$  Zenon Alexa Fluor 488 and incubate 5 minutes at room temperature (concentration of this mix is 0.2  $\mu\text{g}/\mu\text{L}$   $\rightarrow$  18  $\mu\text{g}$  antibody / 90  $\mu\text{L}$  total volume).
  - Dilute 1:10  $\rightarrow$  add 810  $\mu\text{L}$  TBS containing 4% BSA for a total volume of 900  $\mu\text{L}$  (final concentration of antibody is 0.02  $\mu\text{g}/\mu\text{L}$ ).
- b. In a Corning 96-well round bottom plate (Corning #3788) add 10  $\mu\text{L}$  of cells and 15  $\mu\text{L}$  of the antibody mix. Mix with a vortexer set at a slow speed.
  - c. Incubate the plate at 4°C for 20 minutes.
  - d. Add 175  $\mu\text{L}$  of TBS.
  - e. Read the plate on the Guava EasyCyte.

## References

Hanson MA, Brooun A, Baker KA, Jaakola VP, Roth C, Chien EY, Alexandrov A, Velasquez J, Davis L, Griffith M, Moy K, Ganser-Pornillos BK, Hua Y, Kuhn P, Ellis S, Yeager M, Stevens RC., "Profiling of membrane protein variants in a baculovirus system by coupling cell-surface detection with small-scale parallel expression.", *Protein Expr Purif.* (2007) ;**56**(1):85-92.

Please send comments, suggestions, and/or questions to Professor Ray Stevens (stevens@scripps.edu)